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(54) Transfer of macromolecules from a chromatographic substrate to an immobilizing matrix.

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EP-A- 0 050 864
EP-A- 0 061 610
EP-A- 0 066 814
US-A- 4 357 311

ANALYTICAL BIOCHEMISTRY, vol. 124, no. 2, August 1982, pages 396-405, Academic Press, New York, US; J.M. GERSHONI et al.: "Electrophoretic transfer of proteins from sodium dodecyl sulfate-polyacrylamide gels to a positively charged membrane filter"

IDEM

- (73)** Proprietor: **YALE UNIVERSITY**
451 College Street
New Haven, CT 06511(US)
- (72)** Inventor: **Gershoni, Jonathan M.**
The Weizmann Inst. of Science
Dpt. of Biophysics
IL-76100 Rehovot Israel(IL)
- (74)** Representative: **Kraus, Walter, Dr. et al**
Patentanwälte Kraus, Welsert & Partner
Thomas-Wimmer-Ring 15
D-80539 München (DE)

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Description

This invention relates to a solid phase assay system wherein a macromolecule is immobilized in the solid phase on a surface of a chromatographic substrate using a charge modified microporous membrane.

Smithies, Zone Electrophoresis in Starch Gels: Group Variations in the Serum Proteins of Normal Human Adults, *Biochem. J.* 61: 629-641 (1955), showed that starch gel could serve as a molecular sieve through which zone electrophoresis of proteins occur. Since then, there have been constant innovations in the technique of gel electrophoresis. The introduction of acrylamide gels, discontinuous buffer systems, the use of sodium dodecyl sulfate (SDS) to disaggregate protein complexes to be resolved on gels, and the eventual combined use of SDS in discontinuous buffer systems for polyacrylamide gel electrophoresis have been major contributions to the development of one of the most widely used analytical and preparative tools of modern biology.

The main objective of these techniques has been to visually demonstrate the homogeneity or complexity of a protein preparation by following the appearance or disappearance of a particular "band" throughout a given experimental procedure. One-dimensional gels were found to be adequate, provided only relatively simple protein samples such as viruses, bacteriophages, erythrocyte ghost membranes, etc., were being analyzed. More complex systems demanded greater resolving power and new two-dimensional gel systems were developed. Today, even the thousands of polypeptides which are a part of the more intricate proteinaceous samples can be efficiently resolved.

The task of unequivocally correlating a "band" or "spot" with a recognized function has often been difficult, and this is even more so when the resolution of the proteins depends on their denaturation. Nevertheless, many approaches have been developed which allow the identification of a specific enzyme or antigen or glycoprotein or hormone receptor, etc. in a gel. These techniques rely on the ability to maintain at least one of the following prerequisites: (1) that the polypeptides retain their activity throughout electrophoresis; (2) renaturation of a denatured polypeptide; and (3) covalent crosslinkage of the protein in question to a detectable ligand during electrophoresis. Moreover, the actual processing of the gels entails multiple manipulations and extensive incubations and washing procedures. This is very time consuming and quite often prone to handling accidents such as breakage and tearing of wet gels or cracking during the drying of the gels.

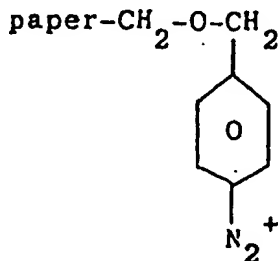
In order to try to overcome some of the problems encountered in analyzing gels, a new approach has evolved. A number of reports have been published demonstrating that the well established approach of "Southern-blotting", i.e. transferring DNA patterns from agarose gels to nitrocellulose membranes, can be applied to protein patterns in polyacrylamide gels. Intact protein patterns are eluted from the gels and are immobilized on a substratum. The substratum is, in turn, subjected to the same type of procedures which have been used on gels for "band" or "spot" identification. However, by transferring electrophoretograms to immobilizing matrices one may benefit from the following advantages: 1) wet immobilizing matrices are pliable and easy to handle; 2) the immobilized proteins are readily and equally accessible to various ligands (since the limitations introduced in gels by differential porosity are obviated); 3) transfer analysis generally calls for small amounts of reagents; 4) processing times (incubations and washings) are significantly reduced; 5) multiple replicas of the gels may be made; 6) transferred patterns may be stored for months prior to their use; 7) protein transfers may undergo multiple analyses. Moreover, the transferred protein patterns are amenable to analyses which would be otherwise extremely difficult or impossible to perform on gels.

The term "blotting" today refers to the process of transferring biological macromolecules such as nucleic acids and proteins from gels to an immobilizing matrix. The term is often used in conjunction with the relevant macromolecule, e.g. protein blotting, DNA blotting and RNA blotting. The resulting matrix containing the transferred immobilized macromolecule is known as a "blot" or "transfer" and can be incubated with a ligand, a procedure which may be referred to as "overlay". Thus, for example, immuno-overlay, lectin overlay or calmodulin overlay refers to the incubation of a blot with an antibody, lectin or calmodulin, respectively.

DNA blotting, a type of nucleic acid blotting, traces its origin to the technique often referred to as a "Southern Transfer" which was developed by Southern, Detection of Specific Sequences among DNA Fragments Separated by Gel Electrophoresis, *J. Mol. Biol.* 98: 503-517 (1975). After the chromatographic separation of the DNA fragments, the DNA is denatured while in the gel and the gel is neutralized. The gel is placed between wicking paper which is in contact with a buffer reservoir, nitrocellulose is placed on top of the gel and dry blotting papers are placed on top of the nitrocellulose. Mass flow of buffer through the gel elutes the DNA which then binds to the nitrocellulose. Thus, the electrophoretically separated DNA fragment pattern is transferred and preserved on the nitrocellulose. Hybridization with a specific labeled nucleic acid

allows detection of the specific complementary fragments bound to the nitrocellulose.

In 1976, it was discovered that single stranded RNA and DNA could be covalently coupled to a cellulose powder substituted with aminobenzyloxymethyl groups which were activated by diazotizing the amine forming diazobenzyloxymethyl (DBM) - cellulose. This filled a gap in hybridization technology since RNA does not bind well to nitrocellulose making a Southern Transfer difficult or impossible. In 1977, Alwine, et al., "Method for Detection of Specific RNAs in Agarose Gels by Transfer to Diazobenzyloxymethyl-Paper and Hybridization with DNA Probes, *Proc. Natl. Acad. Sci. U.S.A.*, 74: 5350-5354, prepared a cellulosic fibrous sheet (i.e. blotting paper) derivatized with diazobenzyloxymethyl groups, termed DBM-paper, viz.



which could be used for transfer of an electrophoretically separated pattern of RNA from an agarose gel in a method similar to a Southern Transfer. Aminophenylthioether paper activated to the diazo form (DPT-paper) has also been used. Both papers covalently and irreversibly couple DNA, RNA and proteins.

DBM-paper and DPT-paper having the disadvantages that they require activation, have limited life, i.e. their activity is labile, having binding capacity only comparable to nitrocellulose, irreversibly couples the macromolecules, thus preventing their subsequent elution, and may present difficulties in resolution due to the texture of the surface.

In an effort to overcome some of the inconveniences of DBM paper, Thomas, (1980) developed a technique to transfer RNA and small DNA fragments to nitrocellulose using high salt concentrations. The binding efficiency of RNA was found to be 80 ug/cm² as compared to 35 ug/cm² for DBM paper. Elution of macromolecules from polyacrylamide gels can be accomplished efficiently by electrophoresis. However, the requirement of high salt concentrations would lead to impractically high currents.

Although first developed in connection with the study of DNA and RNA, it became recognized that the blotting techniques pioneered by Southern could be applied to proteins. The protein transfer techniques were first developed by Renart, *Transfer of Proteins from Gels to Diazobenzyloxymethyl-Paper and Detection with Antisera: A Method for Studying Antibody Specificity and Antigen Structure, Proc. Natl. Acad. Sci. U.S.A.* 76: 3116-3120 (1979), who achieved protein transfer to DBM paper using composite agarose-acrylamide gels in which the acrylamide crosslinking agent was reversible. After electrophoresis, the crosslinking was removed, leaving a low percentage agarose gel from which the proteins transferred easily. Shortly thereafter, a protein blotting procedure was developed by Bowen et al., *The Detection of DNA-Binding Proteins by Protein Blotting, Nuc. Acids Res.* 8: 1-20 (1980), using nitrocellulose for the transfer of DNA binding and other ligand-binding proteins separated on SDS-polyacrylamide gels. In this procedure, transfer was accomplished by diffusion. Towbin, et al. (1979) *Proc. Natl. Acad. Sci. USA* 76: 4350-4354 and Bittner, M. et al. (1980) *Anal. Biochem.* 102: 459-471 had demonstrated that transfer could be accomplished electrophoretically even at low salt concentrations.

In general, protein blotting should be viewed as two sequential events, namely the elution of the polypeptide from the gel and the adsorption of the eluted material to an immobilizing matrix.

Three main driving forces have been exploited for macromolecule elution. One is diffusion. Here, the gel containing the macromolecules to be transferred is sandwiched between two sheets of immobilizing matrix which are in turn sandwiched between foam pads and stainless steel screens. This final assembly is then submerged in two liters of buffer and allowed to sit for 36-48 hours. The result of this incubation is that two identical replica blots are obtained. The efficiency of transfer may reach 75%, but this value must be divided between the two replicas. If the amount of macromolecule adsorbed onto the matrix is sufficient for the intended assays and the long transfer time is not detrimental, blotting by diffusion can be useful.

The second means of macromolecule blotting is essentially based on mass flow of liquid through the gel in the same manner DNA blots are achieved in the traditional procedure described by Southern. The gel is placed in a reservoir of buffer. A matrix is applied to the gel and paper towels are piled onto the matrix. The towels absorb the buffer from the reservoir through the gel and matrix. This movement of fluid serves

as a driving force which elutes the proteins out of the gel which are then trapped in the filter. This technique is less time consuming than diffusion blotting and the efficiency of elution is better. A modification of this approach has been suggested which allows bidirectional blotting. Moreover, the time for efficient elution has been dramatically reduced by applying a vacuum to facilitate the process.

5 The most widely used mode for protein blotting is based on electroeluting the proteins from gels. This is made possible due to the fact that proteins, in contrast to DNA, adsorb the nitrocellulose even in low ionic strength buffers (when other immobilizing matrices are used, e.g. DBM paper, this is no longer a consideration). Therefore, one can electrophorese the proteins out of the gel without generating intolerable currents. It should be noted that the concept of electroelution of macromolecules for blotting was originally described by Arnheim and Southern in 1977 (*Heterogeneity of the Ribosomal Genes in Mice and Men, Cell* 11: 363-370). Numerous apparatus designs have been reported and quite a few are now commercially available. In essence, a wet matrix material is placed on a gel, making sure that no air bubbles are caught within the filter or between the matrix and the gel. The matrix and gel are then sandwiched between supportive porous pads such as "Scotch Brite" scouring pads, foam rubber or layers of wet blotting paper. 15 The assembly is then supported by solid grids (usually nonconductive). It is very important that the gel and matrix are firmly held together. This ensures good transfer and prevents distortion of the protein bands. The supported "gel + matrix sandwich" is inserted into a tank containing "transfer buffer" and placed between two electrodes. The electrodes, which may be tacked to the sides of the tank, are designed so as to generate a homogeneous field over the entire area of the gel which is to be transferred. Continuous 20 conductive sheets can serve as electrodes and theoretically are most appropriate for this purpose. Slabs of graphite and stainless steel plates have been used. However, operating units with such electrodes is usually impractical due to the requirement for excessively high currents. Some apparatus use stainless steel mesh or platinum mesh. An economical, yet efficient, design that seems to work reasonably well is that described by Bittner et al (*Electrophoretic Transfer of Proteins and Nucleic Acids from Slab Gels to Diazobenzoyloxymethyl Cellulose or Nitrocellulose Sheets, Ana. Biochem. 102: 459-471*). Factors that may influence the homogeneity of the field are the distance between the gel and the electrode and the density of the electrode material i.e. the distance between each stretch of wire used. In general, the more electrode material present the lower the electrical resistance of the system. This, therefore, demands higher currents in order to obtain reasonable voltage differences to drive the elution process. The transfer buffers used can 30 be of low ionic strength, such as a phosphate buffer, Tris-borate buffer or Tris-glycine and may or may not contain methanol. Methanol tends to increase the binding capacity of nitrocellulose for protein and stabilize the geometry of the gel being transferred but reduces the elution efficiency of protein from SDS gels and in its presence, electroelution must be carried out for long durations, generally more than 12 hours, in order to obtain efficient transfer of high molecular weight proteins. In the absence of methanol, gels of acrylamide 35 tend to swell and, if allowed to occur during protein transfer, causes the band to be distorted.

The conditions of transferring per se are dependent on the type of gel, the immobilizing matrix and the transfer apparatus used as well as the macromolecules themselves. Nondenaturing gels, SDS gels, lithium dodecyl sulfate containing gels, iso- electrofocusing 2D gels and agarose gels have all been used for protein blotting. It is necessary to determine the electric charge of the protein to be eluted and place the 40 matrix on the appropriate side of the gel. Proteins from SDS - gels, for example, are eluted as anions and therefore the matrix should be placed on the anode side of the gel. The case may be the opposite for nondenaturing gels. As electroelution progresses, electrolytes from the gel are also eluted and contribute to the conductivity of the buffer, resulting in a drop in resistance. Were one to conduct the transfer at constant voltage, the current would increase accordingly and currents in excess of one ampere, and indeed up to 45 five amperes, may develop. High current (one to five ampere) power supplies are now commercially available for electroblotting. Another alternative has been to use a 12V battery charger which seems to be quite adequate. However, as most of the common power supplies used for gel electrophoresis cannot exceed 200 - 250 mA, it has been found advantageous to run at a maximal constant current (i.e. 200 mA) and allow the voltage to gradually drop during the transfer.

50 There are incidences where isolated proteins of low or moderate molecular weight do not elute efficiently from a gel. This can occur in those cases where these proteins fortuitously are at their isoelectric point and have no tendency to migrate in the electric field exerted. In such an instance, other buffer conditions can be employed.

There are numerous immobilizing matrices which are available today. While nitrocellulose is the most 55 widely used material as a matrix, the interaction of a protein with the nitrocellulose is complex and not clearly understood. For example, at pH 8 where protein electroelution is usually performed, nitrocellulose is negatively charged as are the proteins being adsorbed. Hydrophobic effects play a role in this interaction and indeed, protein elution from the matrix is facilitated by non-ionic detergents. Some proteins, especially

those of low molecular weight, bind with low affinities to nitrocellulose and may be lost during transfer or subsequent processing. To prevent such a loss, the transferred polypeptide can be crosslinked to the matrix, thereby covalently stabilizing the protein pattern. The presence of cellulose acetate in nitrocellulose membrane matrices seems to reduce their capacity to bind protein. However, cellulose acetate matrices
 5 have been employed successfully for protein blotting.

In order to remedy some of the disadvantages of nitrocellulose matrices, alternative immobilizing matrices have been proposed. Transfer to DBM paper results in a covalently bound and stable protein pattern. Resolution is slightly lower on this material due to its intrinsic coarseness as compared to membrane matrices. Also, glycine, a commonly used material in transfer buffers, can interfere with DBM
 10 paper protein blotting. There still remains, therefore, a need for an immobilizing matrix which overcomes the disadvantages of the nitrocellulose, DBM paper and other matrices which have been used heretofore. A greater transfer capacity is also desirable.

It is known in the art that membranes may be used in chromatography in general and electrophoresis in particular. See, for example, U.S. Patents:

15 3,808,118 to Golias
 3,829,370 to Bourat
 3,945,926 to Kesting
 3,957,651 to Kesting
 3,989,613 to Gritzner
 20 4,043,895 to Gritzner
 4,111,784 to Dahms
 4,158,683 to Del Campo
 4,204,929 to Bier
 4,243,507 to Martin
 25 4,310,408 to Roe and
 4,311,574 to Ishikawa

Additionally, it is known that polyamide powders may be used to perform chromatographic separations. See, e.g. U.S. Patents:

3,418,158 to Perry and
 30 3,523,350 to Goldberg.

Hiratsuka et al., U.S. Patent 4,128,470, teaches that nylon microporous membranes may be used in electrophoresis and isoelectric focusing as the medium through which chromatography is performed.

New England Nuclear has been marketing an uncharged nylon membrane for use in blotting in its "Gene Screen" electrophoresis product line. This material appears to have equivalent binding capacity for
 35 proteins as nitrocellulose.

Analytical Biochemistry 124, 396 to 405 (1982), discloses a product comprising a chromatographic substrate having on a surface thereof as an immobilising matrix a nylon-66 membrane which has been modified by extensive cationisation. This document does not disclose in words that the nylon membrane is skinless, hydrophilic and microporous, having micropores throughout the membrane, the surfaces of the
 40 micropores being modified by a cationic charge modifying agent which is a reaction product of a polyamine with epichlorohydrin, the reaction product having:

- (i) tertiary amine or quaternary ammonium groups, and
- (ii) epoxide groups along a polyamine chain, the epoxide groups capable of bonding to the membrane.

In this document, the modified membrane is identified as Zeta-bind. Zeta-bind is the same as Zetapor
 45 referred to in the description. However, at the priority date of this application, Zeta-bind would not have been made available to third parties, and therefore, document A does not disclose this invention.

US-A-4 357 311 relates to cellulose ester microporous membranes for preparing solid phase assay systems useful for immobilizing antibodies. According to this reference, cellulose acetate is used and a positively charged material is taken and another filter is added under it. The result is a cellulose acetate
 50 over glass filter. In Example 1 of this reference, it is described how to prepare such a filter. A series of solutions and rinses to activate the filter is used. Then, trichlorotriazine is added to obtain a positive charge. The result is a cellulose acetate positively charged filter.

Thereafter, the filter is incubated for about 40 hours at room temperature (compare column 5 of US-A-4 357 311). The known filter is incubated with a goat antibody against a rabbit antibody. The point is that a
 55 molecule is obtained which has the characteristic of specifically recognizing and binding to any rabbit antibody. It is important to recognize that a filter with a goat antibody stuck to the surface that is coated or applied to the surface thereof is obtained.

It should also be pointed out that the filter according to this reference requires incubation with a second antibody which in turn binds to the first antibody. This point is a critical point for distinguishing the teaching of the prior art from the present application, because with the known filter one has two antibodies. Thereafter, the presence of theophylline in any blood serum may be determined by filtering the blood
 5 through the known filter. The theophylline present will bind to the rabbit antibody.

What is done according to the citation is the opposite of what is done according to this invention, since according to this invention theophylline binds directly to the matrix, and preactivation or incubation of the matrix is not required. According to this invention the matrix can be used directly.

Macromolecule blots or transfers can be "overlaid" with a variety of reagents in much the same manner
 10 that has been developed for gels. Manipulation of filter matrices is less time consuming, more economical with respect to the reagents used and is less exposed to handling accidents. More important, however, is the fact that transferring proteins from gels to matrices in effect eliminates diffusion barriers. Furthermore, denatured polypeptides can sometimes be renatured upon removal of SDS from them and this process is probably much more convenient and effective using blots.

Presently, most of the probes which have been used are macromolecules which specifically bind well to
 15 defined domains of the polypeptides under investigation. Lectins have allowed the detection of glycoproteins and antibodies and the identification of their corresponding antigens. Regardless of what the intended assay may be, vacant areas of the matrix which do not contain protein bands can non-specifically adsorb probes during the overlay process leading to intolerable background. Therefore, the unbound sites
 20 of the matrix must be quenched prior to overlaying the blot. Quenching is most commonly achieved by incubating the blot in high concentrations of bovine serum albumin (BSA) or hemoglobin at 25-60 °C for 1-12 hours. Other materials such as ovalbumin, gelatin and various animal sera have also been used. The temperature, choice of protein and duration of quench depend on the type of matrix material and the probe being used. Non-ionic detergents have also been included in quench or washing buffers to reduce non-
 25 specific binding.

Once the blot has been quenched, it is reacted with the probe. In general, all reactions should be carried out in the presence of quenching protein. The reacted blot is then washed extensively in buffer (which does not have to contain protein). If the probe is itself radioactive or conjugated to an enzyme or
 30 fluorescent tag, the blot can be immediately autoradiographed, reacted with the relevant substrate or visualized in UV light, respectively. If further second or third reagents are necessary to detect the presence of probe-band complexes, then each consecutive reagent is incubated with the blot followed by a wash. The great sensitivity of these overlay techniques has allowed the detection of very low amounts (e.g. 1 ng) of viral antigens in natural fluids. Furthermore, these techniques have also been employed in the analysis of human sera of patient suffering from various immune disorders.

One of the advantages of blots over gels is that they may be reused or subjected to multiple reactions. Once a signal has been obtained and recorded, the blot may be "erased" by removing the probe but retaining the original protein pattern on the matrix. The "erased" filter can be reused for additional overlay
 35 analysis for further characterization of the elements of the gel pattern. "Erasing" can be accomplished by dropping the pH to dissociate antibody-antigen complexes or by denaturing the probe by incubating the blot in urea or SDS. Selective dissociation of probe-band complexes, demonstrating specificity, may also be
 40 achieved. Lectins can be selectively competed off with relevant haptens. Calmodulin can be dissociated from calmodulin binding proteins by removing Ca^{++} from the system. These reactions can still be performed even after the protein blot has been autoradiographed to obtain the initial signal.

Use of protein blots usually has the objectives of demonstrating protein-protein or protein-ligand
 45 interactions or of exploiting the production of an immobilized polypeptide as an intermediate step in immunological or biochemical analyses. Both of these approaches have been exploited in novel usages of protein blots. For example:

a: Analysis of protein-ligand associations.

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DNA-protein and RNA-protein interactions have been analyzed by protein blots. Histone H2 associations with H3 and H4 have also been demonstrated.

The epidermal growth factor receptor was identified by hormone overlaying a transferred membrane pattern. Membranes from human epidermoid carcinoma cells (A-431) were prepared and run on SDS-
 55 polyacrylamide gels. The gels were then electroblotted onto DBM paper, quenched and overlaid with epidermal growth factor and subsequently with radioactively labeled antibody to the hormone. One very predominant signal at 150 KD was detected.

b: Identification of enzyme subunits.

The detection of an inactive enzyme on a protein blot has been demonstrated. Phosphodiesterase I, for example, was boiled in 2% SDS for 5 minutes and run on a SDS-polyacrylamide gel. Protein was blotted
 5 into nitrocellulose filters which were reacted with excess antiphosphodiesterase I. The matrix was then incubated with a crude preparation containing active enzyme. The active enzyme bound via unoccupied sites of the antibody to the inactivated resolved subunit immobilized on the matrix. The matrices were then reacted for enzyme activity and the immunocomplexes were thus detected.

10 c: Affinity purification of monospecific antibodies.

Blots have been used for the purification of monospecific antibodies. Polypeptides are resolved on SDS-polyacrylamide gels and blotted onto DBM or CNBr paper. The matrices were overlaid with serum containing polyclonal antibodies. Then single bands containing antigen-antibody complexes were excised
 15 from the matrices from which the monospecific antibody was eluted by incubating the strip in low pH (2-3) buffer. The eluted probe could then be used for immunocytochemical localization studies.

d: Demonstration of cell-protein interactions.

Protein blots have been used to demonstrate specific whole cell-polypeptide interactions. Human plasma was run on SDS-polyacrylamide gels and transferred to nitrocellulose matrices. Once the matrices were quenched, they were incubated with normal rat kidney cells (NRK cells) which specifically bound to immobilized polypeptides presumably involved in cell attachment. The cells were stained with amino black and were found to locate themselves at two discrete bands. These bands were identified as: fibronectin and
 25 a newly discovered 70 KD entity.

It is an object of this invention to provide a solid phase assay system for use in macromolecule blotting with increased versatility, adsorbence and retention compared to known materials.

It is another object of this invention to provide a solid phase assay system for macromolecule blotting which provides for the more efficient transfer of macromolecules to the matrix and overcomes various
 30 deficiencies of the prior art matrices.

It is a further object of this invention to provide a solid phase assay system which can be used in a method for the electrophoretic transfer of proteins from an electrophoresis gel to an immobilizing matrix which has better adsorbence and retention than prior art material such as nitrocellulose, and does not require high salt concentrations in the transfer buffer.

This invention relates to a solid phase assay system wherein a macromolecule is immobilized in the solid phase on a surface of a chromatographic substrate, characterized by employing as said solid phase a hydrophilic, porous positive charge modified microporous membrane comprising an organic microporous membrane having a charge modifying amount of a cationic charge modifying agent bonded to substantially all of the wetted surfaces of said membrane, which is a reaction product of a polyamine with epichlorohydrin, the reaction product having:
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(i) tertiary amine or quaternary ammonium groups, and

(ii) epoxide groups along a polyamine chain, the epoxide groups capable of bonding to the membrane.

Preferably, the cationic charge modifying agent is a water-soluble organic polymer having a molecular weight greater than about 1,000, wherein each monomer thereof has at least one epoxide group capable of
 45 bonding to the surface of the membrane and at least one tertiary amine or quaternary ammonium group. Most preferably, a portion of the epoxy groups on the organic polymer are bonded to a secondary charge modifying agent which is an aliphatic amine having at least one primary amino or at least two secondary amino groups or an aliphatic amine having at least one secondary amino group and a carboxyl or hydroxyl substituent.

The charge modifying agent can also be an aliphatic amine or polyamine bonded to the membrane through a cross-linking agent which is an aliphatic polyepoxide having a molecular weight of less than about 500.

The preferred microporous membrane is nylon. The charge modified microporous membrane can be employed in the form of a reinforced laminated membrane or preferably a porous reinforcing web
 55 impregnated with a polymeric microporous membrane.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 illustrates the binding of ^{125}I -labeled BSA to a membrane used in this invention and a matrix of the prior art.

5 Figure 1a illustrates the effect of variable concentrations of BSA on the binding of ^{125}I -labeled proteins to IMs.

Figure 1b illustrates the effect of variable concentrations of bovine hemoglobin on the binding of ^{125}I -labeled proteins to IMs.

10 Figure 2 is a quantification of transfer of variable amounts of ^{125}I -labeled BSA to the matrices of Figure 1.

Figure 3 shows the recovery of ^{125}I -labeled BSA on successive layers of the matrices of Figure 1.

Figure 4 is a comparison of BSA recovery on layers of the matrices of Figure 1.

The immobilizing matrix used in this invention is a hydrophilic charge modified microporous membrane comprising an organic microporous membrane having a charge modifying amount of a cationic charge
15 modifying agent bonded to substantially all of the wetted surfaces of the membrane. Microporous membranes and cationic charge modified membranes are well known in the art for the filtration of fluids, e.g. liquids. The cationic charge modified microporous membranes, their preparation and such used in the filtration of fluids are described and claimed in EP-A-0066 814 in the name of Ostreicher et al and EP-A-0050 864 in the name of Barnes et al describes the use of charged modified membrane for the filtration of
20 high purity water (18 megohm-centimeter resistivity) used in the electronics industry; and Ostreicher et al describes the use of charged modified membrane for the filtration of parenteral or body liquids. Additionally, it should be noted that these filter membranes may be reinforced by various means.

Additionally, commercially available nylon microporous filter membranes are available from Pall Corp., Glen Cove, NY under the trademark ULTIPOR N₆₆ and N₆₆ POSIDYNE, the latter being a cationically
25 charged modified filter membrane.

Additionally, AMF Inc., Cuno Div., is selling cationically charged modified nylon microporous filter membrane under the trademark ZETAPOR.

By the use of the term "microporous membrane" as used herein, it is meant a preferably substantially symmetrical and isotropic porous membrane having a pore size of at least 0.05 μm or larger or an initial
30 bubble point (IBP), as that term is used herein, in water of less than 8.22 bar (120 psi). The pore size can be up to about 1.2 μm or an IBP of greater than about 0.685 bar (10 psi). By "symmetrical" it is meant that the pore structure is substantially the same on both sides of the membrane. By the use of the term "isotropic", it is meant the membrane has a uniform pore structure throughout the membrane.

The membrane is hydrophilic. By the use of the term "hydrophilic" in describing the microporous
35 membrane, it is meant a membrane which adsorbs or absorbs water. Generally, such hydrophilicity is produced by a sufficient amount of hydroxyl (-OH), carboxyl (-COOH) amino (-NH₂) and/or similar functional groups on the surface of the membrane. Such groups assist in the adsorption and/or absorption of the water onto the membrane. Such hydrophilicity of the immobilizing matrix is a necessary element of this invention. Hydrophilicity of the membrane per se provides adequate bonding of the charge modifying agent to the
40 microporous membrane.

A preferred microporous membrane is one produced from nylon. The term "nylon" is intended to embrace film forming polyamide resins including copolymers and terpolymers which include the recurring amido grouping.

While, generally, the various nylon or polyamide resins are all copolymers of a diamine and a
45 dicarboxylic acid, or homopolymers of a lactam of an amino acid, they vary widely in crystallinity or solid structure, melting point, and other physical properties. Preferred nylons for use in this invention are copolymers of hexamethylene diamine and adipic acid (nylon 66), copolymers of hexamethylene diamine and sebacic acid (nylon 610), and homopolymers of poly- ϵ -caprolactam (nylon 6).

Alternatively, these preferred polyamide resins have a ratio of methylene (CH₂) to amide (NHCO)
50 groups within the range about 5:1 to about 8:1, most preferably about 5:1 to about 7:1. Nylon 6 and nylon 66 each have a ratio of 6:1, whereas nylon 610 has a ratio of 8:1.

The nylon polymers are available in a wide variety of grades, which vary appreciably with respect to molecular weight, within the range from about 15,000 to about 42,000 and in other characteristics.

The highly preferred species of the units composing the polymer chain is polyhexamethylene ad-
55 ipamide, i.e. nylon 66, and molecular weights in the range above about 30,000 are preferred. Polymers free of additives are generally preferred, but the addition of antioxidants or similar additives may have benefit under some conditions.

The preferred membrane substrates are produced by the method disclosed in U.S. Patent No. 3,876,738 to Marinaccio et al. Another method of producing such membranes is described in EP-A-0 005 536

Additionally, any of the hydrophilic microporous membranes commercially available, for example, Pall Corp.'s ULTIPOR N₆₆®(nylon), Millipore's Durapore (polyvinylidene fluoride), and cellulose acetate/nitrate membranes produced by numerous companies, are suitable for cationic charge modifying for use in accordance with this invention.

The preferred nylon membranes, i.e. described in Marinaccio et al and Pall, are characterized by an isotropic structure, having a high effective surface area and a fine internal microstructure of controlled pore dimensions with narrow pore size distribution and adequate pore volume. For example, a representative 0.22 micrometer rated nylon 66 membrane (polyhexamethylene adipamide) exhibits an initial bubble point (IBP) of about 4.1-4.45 bar (45-50 psig), a foam all over point (FAOP) of about 4.45-4.79 bar (50-55 psig), provides a flow of from 70 to 80 ml/min of water at 1.33 bar (5 psig) (47 mm diameter discs), has a surface area (BET, nitrogen adsorption) of about 13 m²/g and a thickness of about 114.3 to 120.65 μm (4.5 to 4.75 mils).

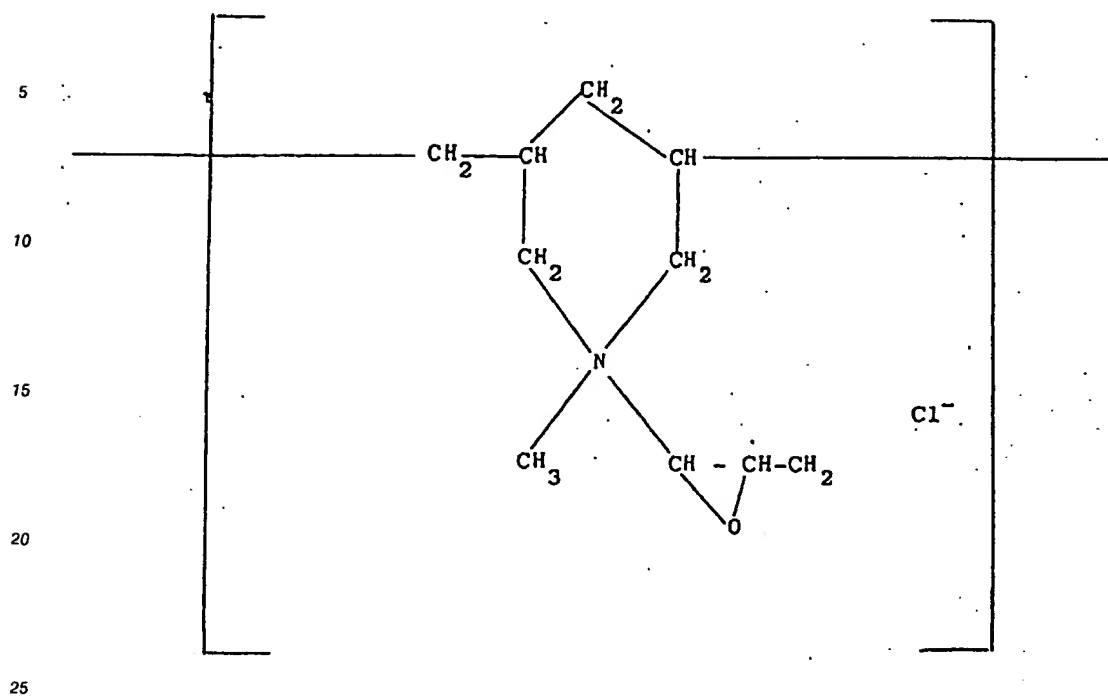
The charge modifying agent is bonded to substantially all of the wetted surface of the microporous membrane. By the use of the term "bonded", it is meant that the charge modifying agent(s) are sufficiently attached to the membrane and/or to each other so that they will not significantly extract under the intended conditions of use. The term "substantially all of the wetted surface" as used herein means that all of the external surface and internal pore surfaces which are wetted by a fluid passing through the membrane or in which the membrane is immersed.

One preferred charge modifying agent which can be used in this invention is described in Ostreicher et al and is a water-soluble organic polymer having a molecular weight greater than about 1,000, wherein the monomer has at least one epoxide substituent capable of bonding to the surface of the membrane and at least one tertiary amine or quaternary ammonium group capable of providing a cationic charge site. Preferably, this charge modifier is a polyamido-polyamine epichlorohydrin cationic resin, in particular, those described in the following U.S. patents:

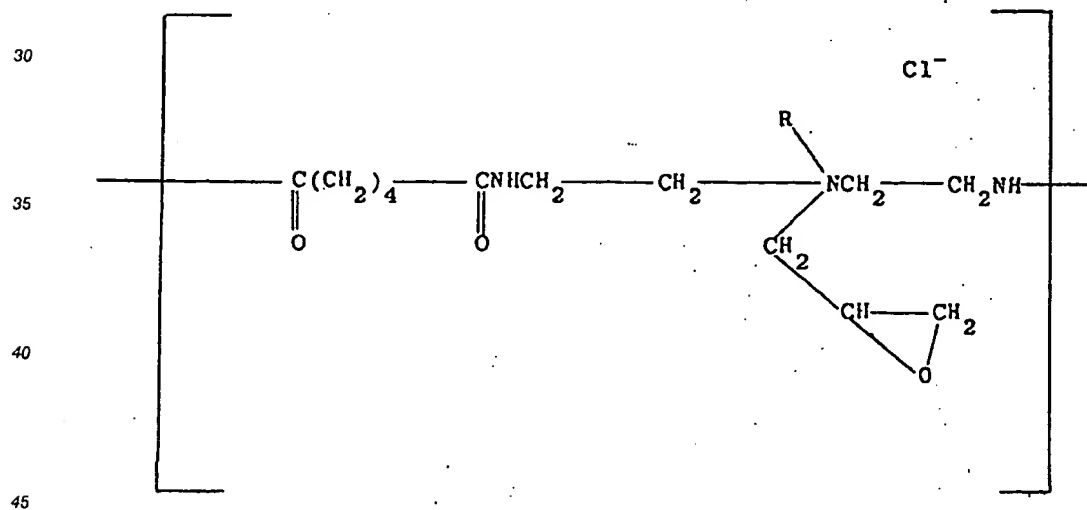
2,926,116 to Keim
 2,926,154 to Keim
 3,224,986 to Butler et al.
 3,311,594 to Earle, Jr.
 3,332,901 to Keim
 3,382,096 to Boardman
 3,761,350 to Munjat et al.

The preferred polyamido-polyamine epichlorohydrin cationic resins are available commercially as Polycup®172, 1884, 2002 or S 2064 (Hercules); Cascamide®Resin pR-420 (Borden); or Nopcobond®35 (Nopco). Most preferably, the polyamido-polyamine epichlorohydrin resin is Hercules R 4308, wherein the charged nitrogen atom forms part of a heterocyclic grouping, and is bonded through a methylene moiety to a depending, reactive epoxide group.

Each monomer group in R 4308 has the general formula:



Polycup 172, 2002 and 1884, on the other hand, have monomer groups of the general formula:



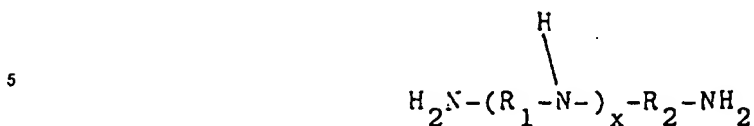
wherein R is methyl or hydrogen (Polycup 172 and 2002, R = H; and Polycup 1884, R = CH₃).

Most preferably, when the charge modifying agent is a water-soluble organic polymer having a molecular weight greater than about 1,000, a secondary charge modifying agent can be used to enhance the cationic charge of the primary charge modifying agent and/or enhance the bonding of the primary charge modifying agent to the microporous surface and/or itself.

The secondary charge modifying agent used in this invention is selected from the group consisting of:

- (i) aliphatic amines having at least one primary amine or at least two secondary amine moieties; and
- (ii) aliphatic amines having at least one secondary amine and a carboxyl or hydroxyl substituent.

Preferably, the secondary charge modifying agent is a polyamine having the formula:



wherein R₁ and R₂ are alkylene of 1 to 4 carbon atoms and x is an integer from 0 to 4. Preferably, R₁ and R₂ are both ethyl.

Preferred polyamines are:

Ethylene diamine H₂N-(CH₂)₂-NH₂

Diethylenetriamine H₂N-(CH₂)₂-NH-(CH₂)₂-NH₂

Triethylenetetramine H₂N-(CH₂-CH₂-NH)₂-CH₂-CH₂-NH₂

15 Tetraethylenepentamine H₂N-(CH₂-CH₂-NH)₃-CH₂-CH₂-NH₂

The highly preferred polyamine is tetraethylene pentamine.

Alternatively, aliphatic amines used in this invention may have at least one secondary amine moiety and a carboxyl or hydroxyl substituent. Exemplary of such aliphatic amines are gamma amino-butyric acid (H₂NCH₂CH₂CH₂COOH) and 2-aminoethanol (H₂NCH₂CH₂OH).

20 The secondary charge modifying agent is bonded to the microporous membrane by bonding to a portion of the epoxide substituents of the polymeric primary charge modifying agent.

The amount of primary and secondary cationic charge modifying agent utilized is an amount sufficient to enhance the electropositive capture potential of the microporous membrane. Such an amount is highly dependent on the specific charge modifying agents utilized.

25 Broadly, the foregoing primary and secondary cationically charge modifying agents are bonded to a hydrophilic organic polymeric microporous membrane, e.g. nylon, by treating the membrane with a charge modifying amount of the primary cationic charge modifying agent and then with the secondary cationically charge modifying agent. Preferably, the process comprises (a) contacting the membrane with an aqueous solution of the primary cationic charge modifying agent and (b) contacting the membrane with an aqueous solution of the secondary charge modifying agent. The contacting steps may be performed in any order, i.e. step (a) prior to step (b) or vice-versa. It is preferred, however, for optimum (minimum) extractables to first contact the membrane with an aqueous solution of the primary cationic charge modifying agent and then subsequently contact the so treated membrane with the aqueous solution of the secondary charge modifying agent.

35 In another embodiment of the present invention, the foregoing secondary charge modifying agent can be used as the charge modifying agent provided it is bonded to the microporous membrane structure through an aliphatic polyepoxide crosslinking agent having a molecular weight of less than about 500. Preferably, the polyepoxide is a di- or tri- epoxide having a molecular weight of from about 146 to about 300. Such polyepoxides have viscosities (undiluted) of less than about 200 centipoises at 25 °C. Due to the necessity of the epoxide to act as a crosslinking agent, monoepoxides, e.g. glycidyl ethers, are unsuitable. Similarly, it is theorized that a polyepoxide offering greater than three epoxy groups offers no benefit and in fact may limit the coupling reactions of the polyepoxide by steric hindrance. Additionally, the presence of unreacted epoxide groups in the cationically charge modified microporous membrane may be undesirable in the finished product.

45 Highly preferred polyepoxides have the formula:



wherein R is an alkyl of 1 to 6 carbon atoms and n is from 2 to 3. the limitation that the number of carbon atoms in the non-epoxide portion --(R)-- be less than 6 is so that the polyepoxide will be soluble in water or ethanol-water mixtures, e.g. up to 20% ethanol. While higher carbon content materials are functionally suitable, their application would involve the use of polar organic solvents with resulting problems in toxicity, flammability and vapor emissions.

The aliphatic amino polyamine charge modifying agent can be bonded to the microporous membrane by (a) contacting the membrane with an aqueous solution of the cationic charge modifying agent and (b)

contacting the membrane with an aqueous solution of the polyepoxide crosslinking agent. The contacting steps may be performed in any order, i.e. step (a) prior to step (b) or vice-versa. Such contacting steps also include contacting the membrane with an aqueous solution of a mixture of the charge modifying agent and the polyepoxide crosslinking agent. It is preferred, however, for optimum (minimum) flushout times to first
 5 contact the membrane with an aqueous solution of the cationic charge modifying agent and then subsequently contact the so treated membrane with the aqueous solution of the polyepoxide crosslinking agent. For maximizing charge modification, however, it is preferred to contact the membrane with an aqueous solution of a mixture of the charge modifying agent and the polyepoxide crosslinking agent.

After the microporous membrane has been contacted with the aqueous solutions, it is then dried and
 10 cured, preferably in a restrained condition, to prevent shrinkage.

Drying of the membrane under restraint is described in the assignee's U.S. Patent 3 769 968.

Generally, any suitable restraining technique may be used while drying, such as winding the membrane tightly about a drying surface, e.g. a drum. Bi-axial control is preferred and tensioning the membrane on a stretching frame is considered the most preferred. Preferably, the restraint imposed affects a reduction in
 15 dimensions.

Final drying and curing temperatures should be to dry and cure the treated membranes, preferably from about 120°C. to 140°C. for minimization of drying times without embrittlement or other detrimental effects to the membrane.

The completed membrane may be rolled and stored for use under ambient conditions.

20 The charge modified microporous membrane can also be employed in the form of a reinforced and/or laminated membrane, preferably one in which a porous reinforcing web is impregnated with a polymeric microporous membrane which is subsequently cationically charged. A preferred reinforced laminated membrane and a porous reinforcing web are described in the aforementioned US-A 3 853 977.

Such an impregnated web is preferably produced by casting a sufficient amount of a casting solution
 25 onto the porous reinforcing web to provide a web having a coating solution thereon which is then contacted with a quenching bath.

The reinforcing web is a porous material which is preferably wettable by the casting solution to maximize impregnation of the casting solution during casting and become firmly attached to the web during precipitation of the polymeric membrane, e.g. nylon. It is not essential, however, that the web be wettable
 30 by the casting solution. If the web is not wettable, the casting solution coating will be largely confined to the surface of the web but is nonetheless adherent thereto due to impregnation of the solution into the web and adhesion of the membrane to the web.

Such wettable and nonwettable reinforcing webs can, for example, be made of nonwoven textiles and cloth, as well as netting of various types, including extruded plastic filament netting, papers and similar
 35 materials. Reinforcing webs which are non-wettable by the casting solution may be fine-pored non-woven webs made from fibers, such as polypropylene or polyethylene. Suitable wettable reinforcing webs include polyesters, as nonwoven fibrous webs or woven webs, using monofilaments or multifilament yarn, the monofilaments being preferred in terms of open structure and lower pressure drops; polyamide fiber woven webs, woven and nonwoven webs of aromatic polyamides, and other relatively polar fibrous products such
 40 as cellulose, regenerated cellulose, cellulose esters, cellulose ethers, glass fiber and similar materials. Cellulosic and synthetic fiber filter papers may also be used as the reinforcing web as well as perforated plastic sheets and open mesh expanded plastics. If the substrate is relatively coarse or in a very open weave structure, even if the fibers are not substantially wetted by the resin solution, the substrate may nonetheless be impregnated by the membrane material. Thus, such non-wettable materials such as
 45 polypropylene and polyethylene can be impregnated by the membrane if they have a sufficiently open structure.

A preferred manner of making the impregnated reinforcing web is by casting a sufficient amount of the casting solution onto the porous reinforcing web to form a web having a coating solution thereon. This coating solution is then calendered, i.e. pressed, preferably by rollers, into the web under conditions of
 50 temperature, pressure and time sufficient to reduce the viscosity of the coating solution sufficiently to ensure enhanced penetration of the coating solution into the web and to remove substantially all entrapped air therefrom to thus form a coated web. Such conditions of temperature, pressure and time are highly dependent on the type reinforcing web utilized, the casting solution, type rollers, etc. Such conditions can be readily determined by one skilled in the art by noting the penetration of the solution into the web, and
 55 pin holes and bubbles in the final coating. The thus coated web is then subsequently treated by casting a sufficient amount of casting solution thereon to form a coated web having an additional coating solution thereon. This so coated web is then quenched in a quenching bath to form the impregnated web to which the outer membranes are then subsequently laminated.

After formation of the reinforced membrane, it can be treated as described hereinabove to produce the cationically charge modified microporous membrane used in this invention.

The cationically charge modified microporous membranes are employed as an immobilizing matrix in macromolecule blotting in the same manner that nitrocellulose and DBM paper has been used heretofore. However, the charge modified microporous membranes of the present invention have a much greater capacity than the immobilizing matrices used heretofore. For example, it has been found that a charge modified microporous membrane of the present invention has a capacity for binding protein of at least 480 ug/cm² while nitrocellulose has a capacity of about 80 ug/cm². It is also not essential to use methanol in the transfer buffer with the immobilizing matrix used in the present invention as is generally the case when nitrocellulose is used. Moreover, nitrocellulose immobilizing matrices, when used in connection with DNA, require high salt concentrations in the buffer; as a result, high current flows of up to 5 amperes are required and the heat generated adversely affects the DNA. The high salt concentrations are not required when using the present charge modified microporous membranes and therefore electrophoresis currents which are in the milliampere range can be employed.

The overlay of the blots (or transferred electrophoretograms) can be carried out in the same manner with the immobilized matrix used in the present invention as they have been done in the past with other immobilizing matrices. As in the case of other matrices, the residual potential binding sites on the matrix must be quenched in order to minimize non-specific background. The high affinity of the charge modified microporous membrane used in this invention for proteins means that the normal quenching procedures may not be sufficient. It has been found that the charge modified microporous membranes used in the present invention can be effectively quenched by incubation in 10% bovine serum albumin (BSA) in phosphate buffered saline (PBS) overnight at 45°-50° C. Hemoglobin (1% in PBS at 45°-50° C) was also found to be effective for quenching the transfers.

While the invention is particularly useful in connection with electrophoretic transfer from chromatographic substrate to matrix, it is also applicable to blotting techniques in general, such as transfer by convection, i.e. mass flow of fluid as a driving force for elution of the macromolecules from the chromatographic substrate to the matrix, and transfer by diffusion.

It is contemplated in accordance with this invention that the cationically charged microporous membrane may be used as an immobilizing matrix for macromolecules derived from not only polyacrylamide or agarose gels but also other chromatographic substrates such as products of thin layer chromatography or high voltage paper electrophoresis or from solutions containing these macromolecules, i.e. directly spotting and to be used for solid phase assays.

Having now generally described this invention, the same will become better understood with reference to certain specific examples, which are included herein for the purpose of illustration only and are not intended to be limiting of the invention.

EXAMPLE I

A.

Preparation of a Microporous Membrane

A representative nylon 66 membrane of 0.22 μ m nominal rating, having a nominal surface area of about 13m²/g, an Initial Bubble Point of about 3.22 bar (47 psi), and a Foam-All-Over-Point of about 3.56 bar (52 psi) was prepared by the method of Marinaccio et al, U.S. Patent 3,876,738, utilizing a dope composition of 16 percent by weight nylon 66 (Monsanto Vydne 66B), 7.1% methanol and 76.9% formic acid, a quench bath composition of 25% methanol, 75% water by volume (regenerated as required by the method of Knight et al, U.S. Patent 3,928,517), a casting speed of 61 cm/min (24 inches/minute) and a quench bath temperature of 20° C. The membrane was cast just under the surface of the quench bath by application to a casting drum rotating in the bath 228.6 - 254 μ m (9 - 10 mils) as cast wet, to obtain 114.3 - 139.7 μ m (4.5 - 5.5 mils) dry and allowed to separate from the drum about 90° of arc from the point of application, the self-supporting membrane forming a shallow catenary to takeup. A portion of the uniform opaque film was dried (in restrained condition to resist shrinkage) in a forced air oven at 80-90° C. for 30 minutes.

B.

Preparation of Charge Modified Microporous Membrane

1. Membrane samples (dried and undried) were dipped in a bath of Hercules 1884 polyamido-polyamine epichlorohydrin resin (4% solids by weight), and allowed to attain adsorption equilibrium. The treated membrane samples were washed to remove excess resin and dried in restrained condition on a drum at a temperature of 110° C. for a period of about 3 minutes.

The treated membrane samples were compared for flow and bubble point characteristics as follows, and found to be essentially identical for treated and untreated samples, evidencing retention of pore and surface geometry. The results are set forth in Table I.

TABLE I

	Control (No Treatment)	Undried Membrane	Dried Membrane
Thickness / μm (mils)	107.95 (4.25)	116.33 (4.58)	122.68 (4.83)
Initial Bubble Point/bar (psi)	2.99 (43.7)	3.06 (44.7)	3.06 (44.7)
Foam-All-Over-Point/bar (psi)	3.77 (55.0)	3.70 (54.0)	3.75 (54.7)
Thickness Normalized Flow Rate / $\text{cc} \cdot \mu\text{m}/\text{min}$ $\cdot \text{cm}^2 \cdot \text{bar}$ ($\text{cc} \cdot \text{mil}/\text{min} \cdot \text{cm}^2 \cdot \text{psi}$)	263.27 (7.1)	266.98 (7.2)	259.56 (7.0)
BET, N_2 adsorption	13.12	-	13.5

The Foam-All-Over-Point (FOAP) is determined by establishing the Initial Bubble Point (IBP) pursuant to ASTM D-2499-66T and then increasing the air pressure until the air flow through the wetted membrane sample, as measured by a flow meter in the line between the regulator and the sample holder, reaches 100 cc/min. FOAP is directly proportional to the mean pore diameter of the sample membrane.

Table I shows, in terms of the morphological and hydrodynamic parameters that control mechanical sieving, the foregoing characteristics of the treated membranes were essentially identical with the untreated nylon membrane.

2. Similar characterizations were conducted on another membrane sample, similarly prepared, but treated with 2% Hercules R 4308 resin (a free radical polymerized resin based upon diallyl nitrogen-containing materials, reacted with epichlorohydrin) in a bath adjusted to pH 10.5, overcoated with 0.1% tetraethylene pentamine, dried, cured, washed and redried. The results are set forth in Table II.

TABLE II

	Tensile Strength/bar (psi)	Control (No treatment)	Treated Membrane
Wet		36.17 (528)	43.50 (635)
Dry		58.91 (860)	65.76 (960)
Elongation (%)			
Wet		140	100
Dry		95	40

Surface area of the treated and untreated membranes remained essentially unchanged; tensile strength increased with treatment with some loss in elongation. The treated sheet was more flexible; creasing of the untreated sheet resulted in cracking and splitting.

EXAMPLE II

Two layers of wet microporous membrane, made as in Example IA, were laminated together and dried to 20-25% moisture. It has been found that membrane in such a wet, swollen condition absorbs charge modifying agents more efficiently than bone dry membrane.

The double layer of membrane was then introduced into a 1.25% by weight solution of Hercules R 4308. The pH of the bath was 10.5.

This bath was produced by diluting 17.25 kg (38 lbs.) of Hercules R 4308 resin from its initial 20% by weight concentration to 5%. Five normal (5N) sodium hydroxide solution was then added to raise the pH to 10.5. The solution was then diluted with D.I. water having greater than 150,000 ohm-cm resistivity in a ratio (volume) 2.5:1. The total volume of bath solution was 227.4 l (60 gallons).

The membrane entered the bath of Hercules R 4308 at an angle of 30° from the horizontal to prevent bubble entrapment in the membrane which can prevent the charge modifying agent from diffusing into the membrane. The membrane was treated in this bath at a speed of 76.2 cm/min (2.5 feet/min) for a length of 121.9 cm (4 feet)

Upon exiting this bath, the membrane was wiped on the bottom surface to remove excess water. A three minute air soak with cool air movement was used before the membrane entered the secondary charge modifying agent bath.

This bath was produced by adding 0.023% tetraethylene pentamine by weight or 0.0513 kg (0.113 lbs.) to 227 liters (60 gallons) of D.I. water (at least 150,000 ohm - cm resistivity). The pH was about 9. The immersion conditions are identical to the first bath of primary charge modifying agent. The membrane was then wrapped around a take-up roll.

The take-up roll of wet membrane was stored for at least 3 hours. The roll was then dried at 121°C (250°F) for 3 minutes to complete the reaction of the charge modifying agents.

The membrane was then washed in a subsequent operation and checked for extraction levels.

EXAMPLE III

Microporous nylon membrane prepared in accordance with Example IA was treated with a Hercules R 4308 primary charge modifying agent (pH of bath adjusted to 10 with sodium hydroxide) and, where indicated, with a polyamine secondary charge modifying agent.

Flow characteristics of the respective membranes showed little or no differentiation, as set forth in Table III:

TABLE III

Treatment			Membrane Characteristics		
Charge Modifier			IBP bar (psi)	FAOP bar (psi)	Thickness Normalized Flow*/cm ² ·μm/min·cm ² bar
Primary	Secondary	Sequence			(cc. ml. / min cm ² psi)
None	None	-	3.34 (48.7)	3.56 (52.0)	250.66 (6.76)
2.0%	0.133% Anquamine**	Primary First	3.24 (47.3)	3.58 (52.3)	250.66 (6.76)
2.0%	0.133% Anquamine	Secondary First	3.11 (45.3)	3.45 (50.3)	250.90 (6.45)
2.0%	0.133% Anquamine	Mixed	3.34 (48.7)	3.51 (51.3)	224.33 (6.05)
2.0%	None	-	3.20 (46.7)	3.49 (51.0)	268.83 (7.25)
2.0%	0.03% Tetraethylene Pentamine	Primary First	3.39 (49.5)	3.70 (54.0)	215.06 (5.80)

*Flow rate (cc) x thickness / μm (mil)
 $(\Delta P \text{ bar (psid)} \times \text{Area (cm}^2\text{)})$

an empirically derived relation to normalize data for
thickness variations.

**Anquamine - 100, a low molecular weight (under 10,000)
cationic polyamide adduct evidencing secondary amine
functionality by comparative UV spectroanalysis, supplied
by Pacific Anchor Chemical Corp.

EXAMPLE IV

In order to compare performance of different primary charge modifiers, particularly polyamide-polyamine epichlorohydrin resin candidates and to optimize application levels and pH conditions, the following tests were conducted, utilizing Hercules resins R 4308, Polycup 172 (pH 4.7 as supplied) and Polycup 2002 (27% solids, pH 3.0 as supplied). The results are set forth in Table IV:

TABLE IV

Primary Charge Modifier	Bath pH	IBP	FAOP
1% R 4308	10.3	47	50
2% R 4308	10.3	45	49
3% R 4308	10.3	44	51
1% 172	11.0	46	50
1% 172	4.9	47	50
2% 172	11.0	47	52
2% R 4308	11.0	47	52
2% 2002	11.0	49	52
Control	-	46	50

EXAMPLE V

A washed and stretch dried nylon microporous membrane prepared in accordance with Example I A. was immersed in a two weight percent solid solution of 1, 4-butanediol diglycid 1 ether solution prepared with an 80-20 mixture of 18 megohm-cm D.I. water in high purity ethanol and having a pH of 6.1-6.4. The membrane was removed from the solution and allowed to drain for about one minute. The membrane was then immersed in a 0.5 weight percent solid solution of tetraethylene pentamine prepared with 18 megohm-cm D.I. water having a pH of 11.2-11.4. The membrane was removed from the solution, allowed to drain for about one minute and then stretch dried at 130 °C. for 5 minutes.

EXAMPLE VI

A nylon impregnated web with a microporous membrane having a nominal pore size rating of about 0.65 μm was produced with a reinforcing web of du Pont Corporation's Reemay 2250 polyester, spun bonded non-woven using a casting solution of 16 weight percent nylon 66, 78.04 weight percent formic acid and 5.96 weight percent methanol. A first outer microporous membrane was brought in contact with the impregnated web to provide a soaking wet contact line at the union of the two layers and a second outer microporous membrane was laid onto the opposite surface of the impregnated web in a similar manner. The three-layer laminated membrane was dried on a Teflon coated steel drum equipped with edge restraining belts on both sides of the laminated membrane and infrared radiant heaters spaced at intervals over the drum circumference. Thereafter, the reinforced laminate was treated with a cationic charge modifier pursuant to Example I B.

EXAMPLE VII

A.

Materials and Methods

Preparation of protein samples:

Erythrocyte ghosts were prepared as described in Fairbanks, G., Steck, T.L. and Wallach, D.F.H. (1971) Biochemistry 10: 2606-2617 from blood accumulated in the pleural cavity upon cardiac puncture of anesthetized CD₁ mice. Bovine brain cortex homogenates were prepared as described in DeCamilli, P., Ueda, T., Bloom, F.E., Battenerg, E., and Greengard, P. (1979) Proc. Natl. Acad. Sci. U.S.A.: 76, 5977-5981. The radioiodination of protein standards, protein A, and concanavalin A was performed with 1,3,4,6-tetrachloro-3a,6a-diphenylglycoluril as described in Fraker, P.J., and Speck, J.C. (1978) Biochem. Biophys. Res. Commun.: 80, 849-857, except that the proteins were dissolved in PBS (1 mg/ml). Free ¹²⁵I was separated from the ¹²⁵I-labeled proteins on either a 5-ml Bio-Gel P-2 column or on a 0.4-ml AG-1-X8 column. The specific activities routinely obtained ranged from 10⁶ to 10⁷ cpm ug⁻¹ protein.

Polyacrylamide gel electrophoresis:

Protein samples were solubilized in buffer containing (final concentrations) 2% (w/v) SDS, 2% (v/v) β -mercaptoethanol, 10% (v/v) glycerol, 0.1% (w/v) bromophenol blue, and 100 mM Tris-HCl; pH 6.8. The samples were boiled in this mixture for 3 min. and then resolved, using Laemmli's system, [Laemmli, U.K. (1970) *Nature (London)*: 227, 680-685] on 10% polyacrylamide slab gels.

Electrophoretic transfer:

Upon completion of electrophoresis, the gel (or portions of it) was placed on a wet Scotch-Brite pad and its surface was rinsed with cool (8-10 °C) transfer buffer (15.6 mM Tris-120 mM glycine, pH 8.3, with or without 20% (v/v) methanol). Tris (41 mM)-boric acid (40 mM), pH 8.3, was also tested. No significant differences were detected between the two buffer systems. The cationically modified microporous membrane of Example II (hereinafter termed "CMMM") or nitrocellulose (hereinafter both are generically referred to as immobilizing matrices "IM") were then wetted by floating it on transfer buffer and placed on the gel, making sure that no air bubbles were caught within the IM or between the latter and the gel. The IM was then covered by a second Scotch-Brite pad. A number of such gel-IM assemblies separated by Scotch-Brite pads can be mounted in sequence for simultaneous transfers. The assembly was placed between plastic grids, which were then snugly inserted -- with the IM toward the anode -- into a Plexiglas tank containing 4 liters of cool transfer buffer. Platinum electrodes in a configuration similar to that described by Bittner et al., *Anal. Biochem.* (1980): 102, 459-471 were secured to the wide walls of the tank. With an 8-cm distance between the anode and the cathode, the electric field generated during electrophoretic transfer appeared to be homogeneous. During transfer, the buffer composition changed as salts were eluted from the gels, and thus the current increased when the voltage was kept constant. Because a standard power supply was used (Buchler Model 3-1500), it was sometimes found more practical to transfer at constant current (200 mA) and let the voltage gradually decrease. In a typical two hour transfer, the voltage dropped, for instance, from 42 to 30 V. The voltage change could be avoided by prior equilibration of the gels with transfer buffer. This procedure could also prevent gel swelling during transfer, a common occurrence at acrylamide concentrations \geq 10%. The IMs could be used immediately after transfer, or dried and stored between sheets of Whatman 3MM chromatography paper. In experiments in which transfer of 125 I-labeled protein was quantitated, both gels and IMs were autoradiographed post-transfer using Kodak XAR-5 film and a DuPont Cronex Lightning Plus intensifying screen at -70 °C. The radioactive bands were excised from gels and IMs and counted in a Beckman Biogamma II counter.

Treatment of transfers with antibodies and lectins:

To prevent nonspecific background binding, the IMs must be quenched. For nitrocellulose, it was sufficient to incubate the filters at room temperature for one hour in 10 mM PBS, pH 7.4, containing either 2% (w/v) BSA or 1% (w/v) hemoglobin. Incubation of CMMM IMs in phosphate buffered saline (PBS), containing either 10% BSA or 1% hemoglobin, for 12 hours at 45-50 °C. was found satisfactory for quenching. The quenched IMs were reacted with the relevant ligands (e.g., antibodies, protein A, lectins) for one hour at room temperature in PBS containing either 2% BSA or 1% hemoglobin and then washed at least 5 times in 50 to 100 ml PBS (20 min each wash). All solutions used in the overlay procedure contained sodium azide (0.05% w/v). The washed IMs were autoradiographed at -70 °C. as described above.

B.

Results

50

1. Binding of Macromolecules to IMs:

(a) Binding experiments were performed to compare the adsorbence of 125 I-labeled BSA to the IMs. Squares of IM material were incubated for one hour in solutions of variable concentrations of 125 I-labeled BSA (0.01-5% w/v in PBS). Each sample of the incubation medium contained an equal tracer amount of 125 I-labeled BSA ($177,000 \pm 6100$ cpm ml $^{-1}$; estimated amount 180 ng ml $^{-1}$). After incubation, the IMs were washed 5 times with 5 ml PBS and counted. The counts were converted to bound BSA (μ g. cm $^{-2}$).

The capacity of the CMMM for binding BSA was consistently higher than that of nitrocellulose. This was particularly evident when high concentrations of BSA were used; at 5% BSA, for instance, the IMs bound 480 ug. cm^{-2} and 80 ug. cm^{-2} , respectively. This is shown in Figure 1 in which the CMMM is designated by ● and the nitrocellulose by ○.

(b) 1 cm^2 pieces of CMMM, nitrocellulose, unmodified nylon and the membrane of Example V were incubated in variable concentrations of hemoglobin (0-5% in PBS) or BSA (0-10% in PBS). Each filter was incubated in 1 ml and rotated 2h RT °c. Then 50 μl of a mixture of ^{125}I -labeled proteins was added to each well. [50 l = 5,000 cpm IgA; 5,000 cpm calmodulin; 5,000 cpm BSA] The filters were thus incubated 1 hr. at 25 °c and then washed 5 times in 1 ml PBS and counted. Background (85 cpm) was subtracted from the results and shown in Figure 1a (BSA) and 1b (hemoglobin). Note, for example, at 1% BSA, the signal of bound radioactive protein has been reduced to 98%, 48%, 94% and 77% when nitrocellulose CMMM, uncharged nylon and the Example V membrane, respectively, were used. The nonquenched IMs were then washed for about 2 hours in 1% Triton®X-100, a non-ionic detergent, to determine how much radioactive protein could be removed. It was found that 40% was removed from the CMMM, about 80% from the IM of Example V, 92% from the uncharged membrane and 92% from the nitrocellulose. These results demonstrate the contribution of the charge modification to the interaction of the proteins with the IMs.

(c) Four Squares (2 cm^2) of IMs were spotted with 50,000 cpm of [^{32}P] labeled *in vitro* transcribed human β -globin mRNA (Kole and Weissman, Nucleic Acid Research, 10: 5429 (1982)) and then washed with PBS until the counts in the wash reached background. The amount of labeled mRNA that bound to the filters was then measured. Squares of commercially available nitrocellulose and cellulose acetate bound less than 100 cpm while the two CMMM (0.45 and 0.2 μm in porosity) bound about 30,000 cpm.

2. Electrophoretic Transfer of Proteins from Gels to Filters:

The IMs were tested in an electrophoretic transfer (1 hour at 30 V) of ^{125}I -labeled BSA (~200 ng), using buffer conditions similar to those described by Towbin et al. (i.e., 15.6 mM Tris, 120 mM glycine, 20% (v/v) methanol, pH 8.3). When methanol was omitted from the transfer buffer, elution increased from 30% to >60 and ~50% of the load when CMMM and nitrocellulose were used, respectively. In the absence of methanol, however, ^{125}I -labeled BSA passed through at least five layers of nitrocellulose (on which it was detected in decreasing amounts, e.g., 15 to 8% of the load); whereas most ^{125}I -labeled BSA (>60% of the load) could be retained on the first CMMM filter, with <1% detected on each sequential filter. Similar results were obtained with other protein standards (phosphorylase b, fetuin, ovalbumin, carbonic anhydrase, and soybean trypsin inhibitor). A detailed analysis of a representative experiment in which ^{125}I -labeled BSA was transferred in the absence of methanol is presented in Figs. 2-4.

Fig. 2 shows quantitation of transfer of variable amounts of ^{125}I -labeled BSA to CMMM or nitrocellulose. Duplicates of four increasing concentrations of ^{125}I -labeled BSA (estimated as 150, 300, 514 and 1400 ng) were run on 10% SDS-polyacrylamide gel. After electrophoresis, one series of lanes was transferred to eight sequential layers of CMMM (●, ■, ▲) and the other to 10 sequential layers of nitrocellulose (○, □, △). At the end of the transfer (2 hours in Tris-boric acid, pH 8.3, without methanol, at constant 32 V), the gels and IMs were counted. The amount of BSA eluted from each gel lane (○, ●) was calculated as the difference between radioactivity loaded and radioactivity remaining in the corresponding gel post-transfer. The unaccounted BSA (△, ▲) is the calculated difference between radioactivity eluted from each gel and radioactivity recovered on the corresponding CMMM or nitrocellulose filters (□, ■).

Fig. 3 shows of ^{125}I -labeled BSA on eight CMMM filters (●) or 10 nitrocellulose filters (○) from the maximal load of BSA (1400 ng) used. Arrow 1 indicates the total amount recovered on eight layers of CMMM. Arrow 2 indicates the total amount recovered on 10 layers of nitrocellulose.

Fig. 4 shows comparison of BSA recovery on the first CMMM filter, the first nitrocellulose layer, and 10 successive nitrocellulose layers. The results obtained in the experiment presented in Fig. 2 were normalized to the amount of BSA recovered on all eight layers of CMMM for each BSA concentration. This value is taken as 100% recovery. Note that for the three lower concentrations of BSA, the first CMMM layer (striped bars) adsorbed as much as or more than all 10 layers of nitrocellulose (hatched bars). In all cases the amount of BSA recovered on the first CMMM layer was considerably greater (4 times) than that recovered on the first nitrocellulose filter (stippled bars).

Figures 2, 3 and 4 show that (i) more than 80% of the protein loaded on the gel could be accounted for when one or two layers of CMMM were used (Figs. 2 and 3); (ii) 80% (at low load) to 50% (at high load) of the IM bound BSA was recovered on the first layer of CMMM (Fig. 4); and (iii) the use of more than three layers of CMMM did not seem to improve the extent of recovery (Fig. 3). Similar results were not obtained

even when 10 layers of nitrocellulose were used (Figs. 2-4). Unaccounted ^{125}I -labeled BSA (presumably lost in the buffer) amounted to >25% of the load when nitrocellulose was used and to $\leq 15\%$ in the case of CMMM.

5 3. Retention of Transferred Proteins on Filters:

Once adsorbed to CMMM, protein is very well retained, as demonstrated by the results of the following experiments. The ^{125}I -labeled BSA run on an SDS-polyacrylamide gel was electrophoretically transferred to either CMMM or nitrocellulose. After transfer, the IMs were incubated in one of the following conditions: (i) 10 15 min in 0.5% glutaraldehyde in PBS, (ii) 1 hr. in 25% isopropanol-10% acetic acid, and (iii) 1 hr. in PBS alone. Next the IMs were rinsed a number of times in PBS and subsequently washed overnight in 0.1% Triton X-100 in PBS. The amount of label on each IM was determined after both transfer and detergent wash. In the case of nitrocellulose, it was found that 80% of the ^{125}I -labeled BSA was washed away from unfixed IMs; although results were variable, at least 1.5-2 times more counts could be retained on such IMs 15 after the glutaraldehyde or the acidic-alcohol treatments. When CMMM was used, >65% of the original counts were retained in the absence of any fixation and fixation increased this value to >90%. Furthermore, the retention of protein to CMMM seemed practically unaffected by variation in the pH (ranging from 2.0 to 8.3) of the washing solutions.

20 4. Overlaying of Transfers:

Before one can use a transferred pattern in any overlay technique, residual potential binding sites on the filter must be quenched to minimize nonspecific background. CMMM was quenched effectively when incubated in 10% BSA in PBS overnight at 45-50°C. Lower temperature (e.g. 37°C.), lower concentrations 25 of BSA, or shorter incubations of the filter with the above solution at 50°C. resulted in unacceptably high background in overlays when these probes were used. Hemoglobin (1% in PBS at 45-50°C.) was also found to be effective for quenching CMMM transfers.

Overlays of protein patterns were transferred to CMMM or nitrocellulose with antibodies or lectins. In particular, aliquots of bovine brain cortex homogenates (25 ug each) were resolved on a 10% SDS-polyacrylamide gel and electrophoretically transferred to either CMMM or nitrocellulose over two hours in 30 Tris-glycine buffer at 200 mA. After quenching with BSA, the IMs were overlaid for one hour with dilute (1:300) rabbit serum containing anti-protein I (synapsin), washed and subsequently incubated with ^{125}I -labeled protein A (10^6 cpm total) for one hour, washed again and then autoradiographed. The same procedure was carried out using aliquots of murine erythrocytic ghosts (25 ug each), filters quenched with 35 hemoglobin and ^{125}I -labeled concanavalin A (5×10^5 cpm total) as the probe. The greater binding capacity of CMMM over nitrocellulose rendered these techniques more sensitive when CMMM is used.

The specificity of lectin binding was tested by using appropriate haptens. In the case of the autoradiographs of the ^{125}I -labeled concanavalin A, for instance, the net radioactivity bound to the respective IMs was determined by deducting from the total counts the background counts measured on the lower parts 40 of the lanes. The net signal on CMMM was 1.6 times higher than on nitrocellulose. A first wash in PBS containing 100 mM α -methylglucoside removed 82 and 76% of the signal from CMMM and nitrocellulose, respectively. A second wash in PBS containing 100 mM α -methylmannoside increased the removal to 90% for CMMM and 84% for nitrocellulose.

The acetylcholine receptor has also been analyzed by protein blotting. Electric organ membranes 45 prepared from *Torpedo* were run at 4°C on polyacrylamide gels containing lithium dodecyl sulfate. The electrophoretograms were electroblotted to CMMM, which were then quenched with hemoglobin and overlaid with ^{125}I -labeled α -bungarotoxin. Only the α -subunit of the receptor bound the toxin. This binding could be competed with nonradioactive α -bungarotoxin and with tubocurarine, another acetylcholine antagonist.

50 Various changes and modifications can be made in the process and products of this invention without departing from the spirit and scope thereof. The various embodiments which have been disclosed herein were for the purpose of further illustrating the invention, but were not intended to limit it.

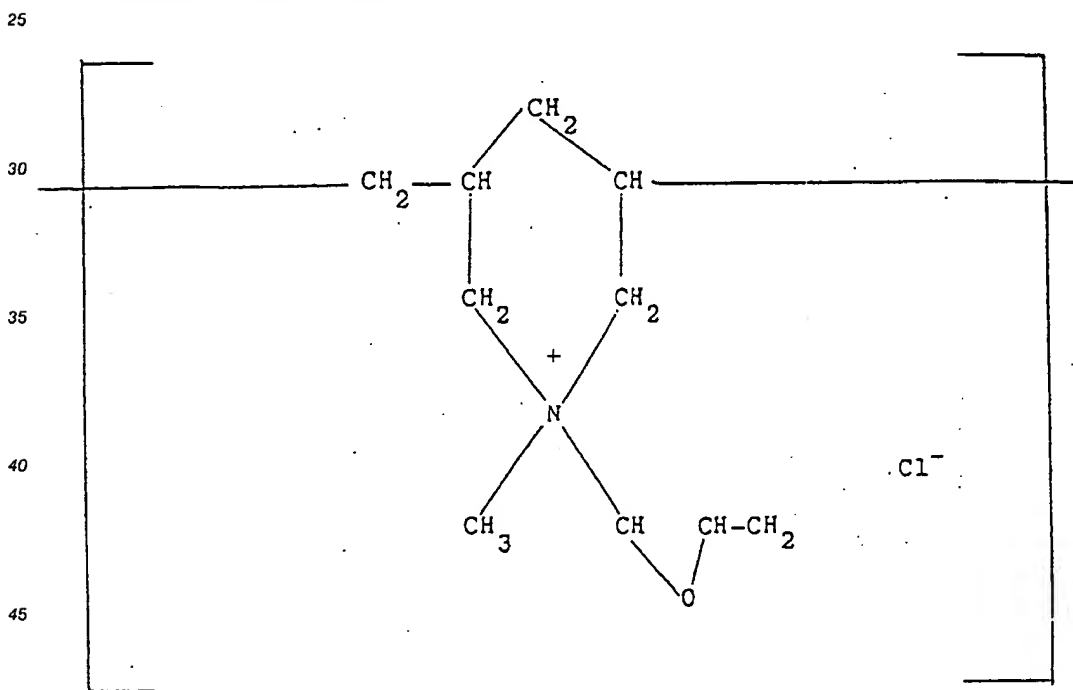
Claims

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1. A solid phase assay system wherein a macromolecule is immobilized in the solid phase on a surface of a chromatographic substrate, characterized by employing as said solid phase a hydrophilic, porous positive charge modified microporous membrane comprising an organic microporous membrane having

a charge modifying amount of a cationic charge modifying agent bonded to substantially all of the wetted surfaces of said membrane, which is a reaction product of a polyamine with epichlorohydrin, the reaction product having:

- 5 (i) tertiary amine or quaternary ammonium groups, and
(ii) epoxide groups along a polyamine chain, the epoxide groups capable of bonding to the membrane.
2. The system of claim 1, characterized in that said organic microporous membrane is hydrophilic.
- 10 3. The system of claim 2, characterized in that the hydrophilic organic polymeric microporous membrane is a cellulose ester.
4. The system of claim 2, characterized in that the hydrophilic organic polymeric microporous membrane is a polyamide.
- 15 5. The system of claim 3, characterized in that the charge modifying agent comprises a water-soluble organic polymer having a molecular weight greater than about 1,000, wherein each monomer thereof has at least one epoxide group capable of bonding to the surface of the membranes and at least one tertiary amine or quaternary ammonium group.
- 20 6. The system of claim 5, characterized in that said water-soluble organic polymer is a polyamido-polyamine epichlorohydrin resin.
7. The system of claim 6, characterized in that said polymer has repeating monomer units of the formula:



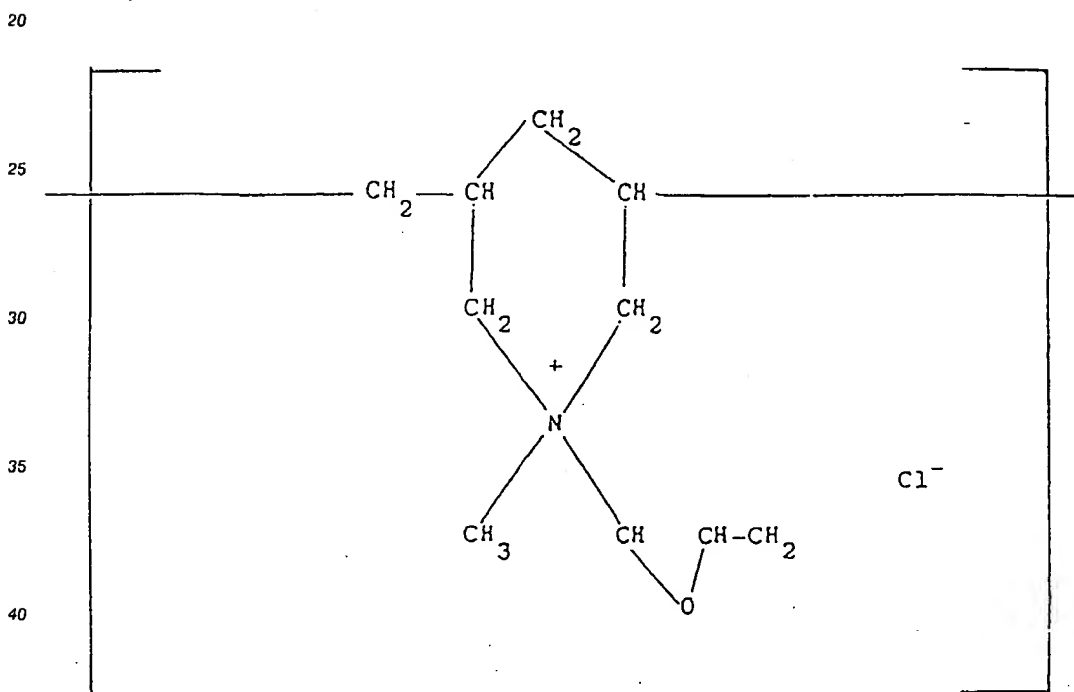
- 50 8. The system of claim 7, characterized in that said membrane is a polyhexamethylene adipamide.
9. The system of claim 5, characterized in that a portion of the epoxy groups of the organic polymer charge modifying agent are bonded to a secondary charge modifying agent selected from the group
- 55 consisting of:
- (i) aliphatic amines having at least one primary amino or at least two secondary amino groups; and
 - (ii) aliphatic amines having at least one secondary amino and a carboxyl or hydroxyl substituent.

10. The system of claim 9, characterized in that said secondary charge modifying agent is tetraethylenepentamine.
11. The system of claim 10, characterized in that said substrate is an electrophoresis gel.
12. The system of claim 1, characterized in that said charge modifying agent is selected from the group consisting of:
- (i) aliphatic amines having at least one primary amino or at least two secondary amino groups; and
 - (ii) aliphatic amines having at least one secondary amino and a carboxyl or hydroxyl substituent, and
- wherein said charge modifying agent is bonded to the microporous membrane structure through an aliphatic polyepoxide cross-linking agent having a molecular weight of less than about 500.
13. The system of claim 12, characterized in that the polyepoxide is 1,4-butanediol diglycidal ether.
14. The system of claim 13, characterized in that said charge modifying agent is tetraethylenepentamine.
15. The system of claim 14, characterized in that said substrate is an electrophoresis gel.
16. The system of claim 4, characterized in that the membrane has a pore size of from about 0.05 to about 1.2 microns.
17. The system of claim 2, characterized in that the charge modified microporous membrane was quenched.
18. A solid phase assay system wherein a macromolecule is immobilized in the solid phase on a surface of a chromatographic substrate, characterized by employing as said solid phase a cationic charge modified microporous membrane comprising a hydrophilic membrane having micropores throughout the membrane, the surfaces of the micropores being modified by a cationic charge modifying agent, which is a reaction product of a polyamine with epichlorohydrin, the reaction product having:
- (i) tertiary amine or quaternary ammonium groups, and
 - (ii) epoxide groups along a polyamine chain, the epoxide groups capable of bonding to the membrane.
19. A solid phase assay system wherein a macromolecule is immobilized in the solid phase on a surface of a chromatographic substrate, characterized by employing as said solid phase a hydrophilic cationic charge modified microporous membrane comprising a substantially isotropic, porous, hydrophilic nylon microporous membrane having a microstructure throughout said membrane, and a charge modifying amount of a cationic charge modifying agent bonded to substantially all of said membrane microstructure without substantial pore size reduction, which is a reaction product of a polyamine with epichlorohydrin, the reaction product having:
- (i) tertiary amine or quaternary ammonium groups, and
 - (ii) epoxide groups along a polyamine chain, the epoxide groups capable of bonding to the membrane.

45 Patentansprüche

1. Assaysystem mit fester Phase, bei dem ein Makromolekül in der festen Phase auf der Oberfläche eines chromatographischen Substrats immobilisiert ist, dadurch **gekennzeichnet**, daß als feste Phase eine hydrophile, poröse durch positive Ladung modifizierte mikroporöse Membran verwendet wird, die eine organische mikroporöse Membran mit einer ladungsmodifizierenden Menge eines kationischen Ladungs-Modifizierungsmittels, das im wesentlichen an die gesamte benetzte Oberfläche der Membran gebunden ist, umfaßt, welches das Reaktionsprodukt aus einem Polyamin mit Epichlorhydrin ist, wobei das Reaktionsprodukt
- (i) tertiäre Amin- oder quaternäre Ammoniumgruppen, und
 - (ii) Epoxidgruppen längs einer Polyaminkette enthält, wobei die Epoxidgruppen an die Membran binden können.

2. System nach Anspruch 1, dadurch **gekennzeichnet**, daß die organische mikroporöse Membran hydrophil ist.
3. System nach Anspruch 2, dadurch **gekennzeichnet**, daß die hydrophile organische polymere mikroporöse Membran ein Celluloseester ist.
4. System nach Anspruch 2, dadurch **gekennzeichnet**, daß die hydrophile organische polymere mikroporöse Membran ein Polyamid ist.
5. System nach Anspruch 3, dadurch **gekennzeichnet**, daß das Ladungs-Modifizierungsmittel ein wasserlösliches organisches Polymeres mit einem Molekulargewicht größer als etwa 1000 enthält, wobei jedes Monomere davon mindestens eine Epoxidgruppe, die in der Lage ist, an die Oberfläche der Membran zu binden, und mindestens eine tertiäre Amin- oder quaternäre Ammoniumgruppe enthält.
6. System nach Anspruch 5, dadurch **gekennzeichnet**, daß das wasserlösliche organische Polymere ein Polyamido-Polyamin-Epichlorhydrin-Harz ist.
7. System nach Anspruch 6, dadurch **gekennzeichnet**, daß das wasserlösliche organische Polymere ein Polymeres ist, das sich wiederholende Monomereinheiten der Formel



- enthält.
8. System nach Anspruch 7, dadurch **gekennzeichnet**, daß die Membran Polyhexamethylenadipamid ist.
9. System nach Anspruch 5, dadurch **gekennzeichnet**, daß ein Teil der Epoxygruppen des Ladungs-Modifizierungsmittels für das organische Polymere an ein sekundäres Ladungs-Modifizierungsmittel gebunden ist, das ausgewählt wird aus der Gruppe, die besteht aus
- (i) aliphatischen Aminen mit mindestens einer primären Amino- oder mindestens zwei sekundären Aminogruppen; und
 - (ii) aliphatischen Aminen mit mindestens einem sekundären Amino- und einem Carboxyl- oder Hydroxylsubstituenten.
10. System nach Anspruch 9, dadurch **gekennzeichnet**, daß das sekundäre Ladungs-Modifizierungsmittel ein Tetraäthylenpentamin ist.

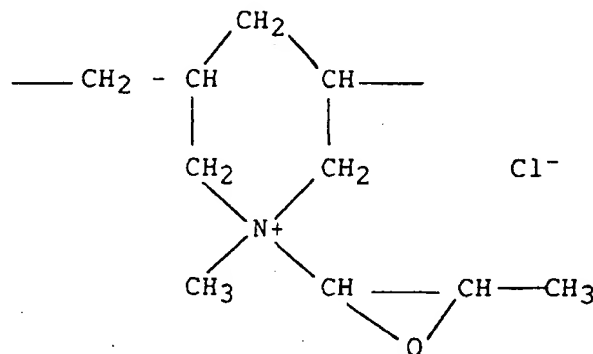
11. System nach Anspruch 10, dadurch **gekennzeichnet**, daß das Substrat ein Elektrophoresegel ist.
12. System nach Anspruch 1, dadurch **gekennzeichnet**, daß das Ladungs-Modifizierungsmittel ausgewählt wird aus der Gruppe, die besteht aus
 - 5 (i) aliphatischen Aminen mit mindestens einer primären Amino- oder mindestens zwei sekundären Aminogruppen; und
 - (ii) aliphatischen Aminen mit mindestens einem sekundären Amino- und einem Carboxyl- oder Hydroxylsubstituenten, und wobei das Ladungs-Modifizierungsmittel an die mikroporöse Membranstruktur über ein aliphatisches Polyepoxid als Vernetzungsmittel mit einem Molekulargewicht unter
 - 10 etwa 500 gebunden ist.
13. System nach Anspruch 12, dadurch **gekennzeichnet**, daß das Polyepoxid 1,4-Butandiol diglycidalether ist.
14. System nach Anspruch 13, dadurch **gekennzeichnet**, daß das Ladungs-Modifizierungsmittel Tetraethylenpentamin ist.
15. System nach Anspruch 14, dadurch **gekennzeichnet**, daß das Substrat ein Elektrophoresegel ist.
16. System nach Anspruch 4, dadurch **gekennzeichnet**, daß die Membran eine Porengröße von etwa 0,05 bis etwa 1,2 μm besitzt.
17. System nach Anspruch 2, dadurch **gekennzeichnet**, daß die ladungsmodifizierte mikroporöse Membran abgeschreckt worden ist.
18. Assaysystem mit fester Phase, bei dem ein Makromolekül in fester Phase auf der Oberfläche eines chromatographischen Substrats immobilisiert worden ist, dadurch **gekennzeichnet**, daß als feste Phase eine mit kationischer Ladung modifizierte mikroporöse Membran verwendet wird, die eine hydrophile Membran mit Mikroporen innerhalb der Membran umfaßt, wobei die Oberflächen der Mikroporen mit einem kationischen Ladungs-Modifizierungsmittel modifiziert worden sind, welches das Reaktionsprodukt eines Polyamins mit Epichlorhydrin ist, wobei das Reaktionsprodukt
 - 25 (i) tertiäre Amin- oder quaternäre Ammoniumgruppen, und
 - (ii) Epoxidgruppen längs einer Polyaminkette enthält, wobei die Epoxidgruppen an die Membran binden können.
19. Assaysystem mit fester Phase, bei dem ein Makromolekül in fester Phase auf der Oberfläche eines chromatographischen Substrats immobilisiert ist, dadurch **gekennzeichnet**, daß als feste Phase eine hydrophile mit kationischer Ladung modifizierte mikroporöse Membran verwendet wird, die eine im wesentlichen isotrop, poröse, hydrophile mikroporöse Nylonmembran mit einer Mikrostruktur innerhalb der Membran und eine ladungsmodifizierende Menge eines kationischen Ladungs-Modifizierungsmittels, das im wesentlichen an die gesamte Membranmikrostruktur ohne wesentliche Porengrößenverringern gebunden ist, umfaßt, welche das Reaktionsprodukt aus einem Polyamin mit Epichlorhydrin ist, wobei das Reaktionsprodukt
 - 30 (i) tertiäre Amin- oder quaternäre Ammoniumgruppen, und
 - (ii) Epoxidgruppen längs einer Polyaminkette enthält, wobei die Epoxidgruppen an die Membran binden können.

Revendications

1. Système d'essai en phase solide dans lequel une macromolécule est immobilisée dans la phase solide à la surface d'un substrat chromatographique, caractérisé en ce que l'on utilise comme phase solide une membrane microporeuse hydrophile modifiée par une charge positive, comprenant une membrane microporeuse organique ayant une quantité modificatrice de charge d'un agent cationique modificateur de charge liée à substantiellement toutes les surfaces humectées de ladite membrane, ledit agent modificateur de charge étant le produit de la réaction d'une polyamine avec l'épichlorhydrine, le produit de la réaction ayant :
 - 50 (i) des groupes amine tertiaire ou ammonium quaternaire, et

(ii) des groupes époxydes le long d'une chaîne de polyamine, les groupes époxydes étant capables de se lier à la membrane.

2. Système selon la revendication 1, caractérisé en ce que ladite membrane microporeuse organique est hydrophile.
3. Système selon la revendication 2, caractérisé en ce que la membrane microporeuse organique hydrophile est un ester de cellulose.
4. Système selon la revendication 2, caractérisé en ce que la membrane microporeuse organique hydrophile est un polyamide.
5. Système selon la revendication 1, caractérisé en ce que l'agent modificateur de charge comprend un polymère organique hydrosoluble ayant une masse moléculaire supérieure à environ 1000, dans lequel chacun des monomères a au moins un groupe époxyde capable de se lier à la surface de la membrane et au moins un groupe amine tertiaire ou ammonium quaternaire.
6. Système selon la revendication 5, caractérisé en ce que ledit polymère organique hydrosoluble est une résine de polyamido-polyamine et d'épichlorhydrine.
7. Système selon la revendication 6, caractérisé en ce que ledit polymère a des motifs monomères de formule :



8. Système selon la revendication 7, caractérisé en ce que ladite membrane est un polyhexaméthylène adipamide.
9. Système selon la revendication 5, caractérisé en ce qu'une partie des groupes époxydes du polymère organique constituant l'agent modificateur de charge sont liés à un agent modificateur de charge secondaire choisi dans le groupe formé par :
 - (i) des amines aliphatiques ayant au moins un groupe amino primaire ou au moins deux groupes amino secondaires ; et
 - (ii) des amines aliphatiques ayant au moins un groupe amino secondaire et un substituant carboxyle ou hydroxyle.
10. Système selon la revendication 9, caractérisé en ce que ledit agent modificateur de charge secondaire est la tétraéthylènepentamine.
11. Système selon la revendication 10, caractérisé en ce que ledit substrat est un gel d'électrophorèse.
12. Système selon la revendication 1, caractérisé en ce que ledit agent modificateur de charge est choisi dans le groupe formé par :
 - (i) des amines aliphatiques ayant au moins un groupe amino primaire ou au moins deux groupes amino secondaires ; et

(ii) des amines aliphatiques ayant au moins un groupe amino secondaire et un substituant carboxyle ou hydroxyle, et dans lequel ledit agent modificateur de charge est lié à la structure de la membrane microporeuse par un agent de réticulation constitué d'un polyépoxyde aliphatique ayant une masse moléculaire inférieure à environ 500.

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13. Système selon la revendication 12, caractérisé en ce que le polyépoxyde est l'éther de 1,4-butanediol diglycidal.

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14. Système selon la revendication 13, caractérisé en ce que ledit agent modificateur de charge est la tétraéthylènepentamine.

15. Système selon la revendication 14, caractérisé en ce que ledit substrat est un gel d'électrophorèse

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16. Système selon la revendication 4, caractérisé en ce que la membrane a une taille de pores d'environ 0,05 à environ 1,2 microns.

17. Système selon la revendication 2, caractérisé en ce que la membrane microporeuse modifiée par une charge a été refroidie brusquement.

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18. Système d'essai en phase solide dans lequel une macromolécule est immobilisée dans la phase solide à la surface d'un substrat chromatographique, caractérisé en ce que l'on utilise comme phase solide une membrane microporeuse modifiée par une charge cationique, comprenant une membrane hydrophile ayant des micropores dans toute sa masse, les surfaces des micropores étant modifiées par un agent modificateur de charge cationique qui est le produit de la réaction d'une polyamine avec l'épichlorhydrine, le produit de la réaction ayant :

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(i) des groupes aminetertiaire ou ammonium quaternaire, et

(ii) des groupes époxydes le long d'une chaîne polyamine, les groupes époxydes étant capables de se lier à la membrane.

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19. Système d'essai en phase solide dans lequel une macromolécule est immobilisée dans la phase solide à la surface d'un substrat chromatographique, caractérisé en ce qu'on utilise comme phase solide une membrane microporeuse hydrophile modifiée par une charge cationique, comprenant une membrane microporeuse de polyamide hydrophile, poreuse, substantiellement isotrope, ayant une microstructure dans toute sa masse, et une quantité modificatrice de charge d'un agent modificateur de charge cationique lié à substantiellement toute ladite microstructure de la membrane sans réduction notable de la taille des pores, l'agent modificateur de charge cationique étant le produit de la réaction d'une polyamine avec l'épichlorhydrine, le produit de la réaction ayant :

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(i) des groupes amine tertiaire ou ammonium quaternaire, et

(ii) des groupes époxydes le long d'une chaîne de polyamine, les groupes époxydes étant capables de se lier à la membrane.

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